

Synthesis of isocorrole and the higher homologues

Jun-ichiro Setsune,* Aki Tsukajima and Junko Watanabe

Department of Chemistry, Faculty of Science, Kobe University, Nada-ku, Kobe 657-8501, Japan

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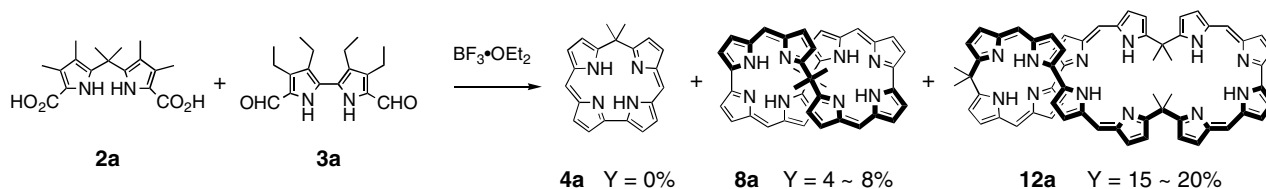
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Abstract—Bis(azafulvene) derivative of *gem*-dimethyldipyrromethane reacted with 2,2'-bipyrrole under neutral conditions without catalyst to give a mixture of expanded isocorroles in ca. 50% total yields. GPC separation gave eleven porphyrinoids containing 4, 8, 12, 16, 20, 24, 28, 32, 36, 40, and 44 units of pyrrole.
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Isocorrole is a tautomeric form of corrole where one of the three *meso*-carbons of the (1.1.1.0)-type tetrapyrrolic macrocycle is sp³-hybridized and two NH protons are inside the cavity,^{1,2} although a (2.0.1.0)-type structural isomer is also called by the same name.³ The isocorrole nucleus has only recently been synthesized by the condensation of two parts of 3,4-diethyl-2-formylpyrrole **1** with *gem*-dimethyl-3,3',4,4'-tetramethyldipyrromethane-5,5'-dicarboxylic acid **2a** followed by the oxidative coupling of the resulting *a,c*-biladiene in the presence of Ni(OAc)₂·4H₂O.² On the other hand, the [2+2]-type condensation of the diacid **2a** and 5,5'-diformyl-3,3',4,4'-tetraethyl-2,2'-bipyrrole **3a** did not give isocorrole **4a** at all, but afforded cyclooctapyrrole **8a** and cyclododecapyrrole **12a** (see Scheme 1).¹ These expanded isocorroles are closely related to the calixphyrins where both sp²- and sp³-hybridized bridging carbons are present in the porphyrin framework.⁴ Thus, more flexible ring structure is expected for the expanded isocorroles in comparison with the expanded porphyrins with complete cycloconjugation. In view of the fact that there has been very little work on the isocorrole deriva-

tives in spite of their importance as a member of porphyrinoids, we have applied our original synthetic method using bis(azafulvene) to the preparation of *gem*-dimethylisocorrole and the higher homologues.⁵

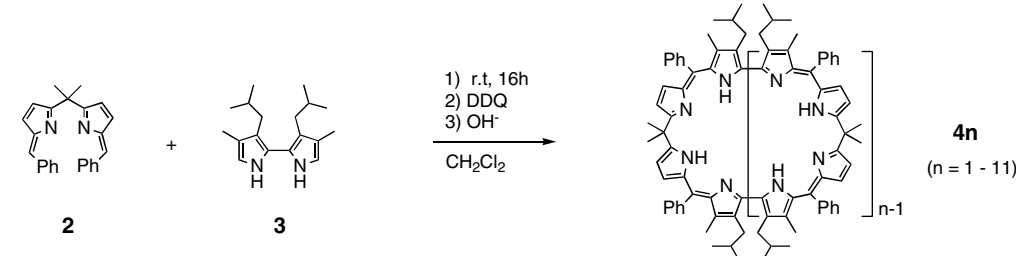
A CH₂Cl₂ solution (40 mL) of *gem*-dimethyl-2,2'-bis(6-phenylazafulvenyl)methane **2**⁶ (0.48 mmol) and 3,3'-di-*iso*-butyl-4,4'-dimethyl-2,2'-bipyrrole **3**⁷ (0.48 mmol) was stirred at room temperature for 16 h under argon. After oxidation with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (abbreviated hereafter as DDQ) (1.44 mmol), the reaction mixture was purified by column chromatography to give isocorrole **4** in 27.8% yield. The higher homologues **8**, **12**, **16**, **20**, and **24** containing 8, 12, 16, 20, and 24 pyrrole units were separated by gel permeation chromatography (GPC) in 9.5%, 5.8%, 3.2%, 2.5%, and 1.5% yield, respectively, as shown in Table 1. When the reactant concentration was increased from 0.012 to 0.027 mol/L, the yield of higher homologues increased in sacrifice of the yield of **4** and the total yield of the macrocycles decreased from 50.5% to 44.2% (Table 1, entry 2). Addition of



Scheme 1. Synthesis of cyclooctapyrrole **8a** and cyclododecapyrrole **12a** by Vogel and co-workers (Ref. 1).

Keywords: Porphyrinoids; Calixphyrin; Macrocycle; Porphyrins; Expanded porphyrins; Corrole.

* Corresponding author. Tel.: +81 78 803 5683; fax: +81 78 803 5770; e-mail: setsunej@kobe-u.ac.jp

Table 1. Synthesis of cyclopolypyrroles **4n**


Entry	Additive ^a	Conc. ^b	Yield (%) of 4n (n = 1–8)								Total
			4	8	12	16	20	24	28	32	
1	None	0.012	27.8	9.5	5.8	3.2	2.5	1.5	tr	—	50.5
2	None	0.027	10.3	10.4	9.4	6.8	3.8	2.3	1.2	tr	44.2
3	Zn(<i>p</i> - <i>t</i> -BuC ₆ H ₄ CO ₂) ₂	0.027	4.5	12.8	9.6	7.6	4.6	2.7	1.6	0.7	44.1

^a Zinc *p*-*tert*-butylbenzoate (2 molar equiv) was added.

^b Concentration of **2** and **3** (mol/L).

2 M equiv of zinc(II) *p*-*tert*-butylbenzoate further changed the products distribution in favor of the higher homologues without affecting the total yield (Table 1, entry 3). Thus, expanded isocorroles **28** and **32** with 28 and 32 pyrrole units were isolated in 1.6% and 0.7% yield, respectively. The first fraction in the GPC was further separated by using polystyrene–polydivinylbenzene gel with a larger pore size to result in the isolation of giant homologues **36**, **40**, and **44** with 36, 40, and 44 pyrrole units in the yield of 0.5–0.1%.⁸ However, the total yield decreased to only 15% at the reactant concentration of 0.080 mol/L.

These expanded isocorroles were identified by ESIMS. The largest homologue, cyclotetratetracontapyrrole **44**, ($M_W = 6807.1$) showed seven signals corresponding to the diprotonated species at 3405.6 mass (3404.6 calcd for C₄₇₃H₅₀₆N₄₄ + 2H⁺) through the octa-protonated species at 852.0 mass (851.9 calcd for C₄₇₃H₅₀₆N₄₄ + 8H⁺) as depicted in Figure 1.

The UV–vis absorption band is red-shifted with increasing the ring size as shown in Figure 2; 420, 541, 538, 557, 574, 582, and 584 nm for **4**, **8**, **12**, **16**, **20**, **24**, and **28**, respectively. The larger homologues **32**, **36**, **40**, and **44** showed virtually the same UV–vis feature with an

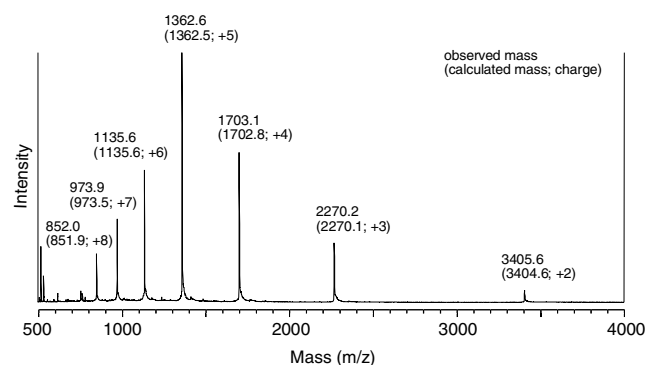


Figure 1. ESIMS spectrum of **44**. The calculated mass and the number of charges are indicated in the parenthesis.

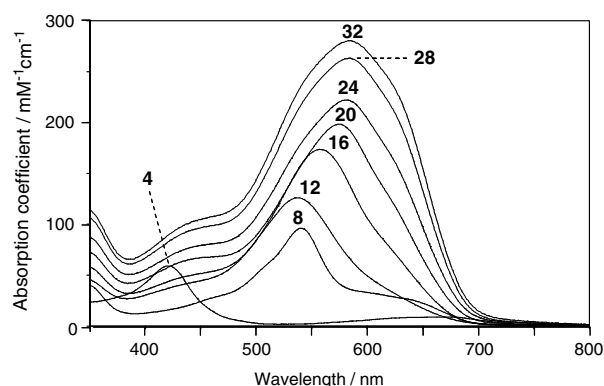


Figure 2. UV–vis spectra of isocorrole **4** and the higher homologues **8**, **12**, **16**, **20**, **24**, **28**, and **32** in CH₂Cl₂.

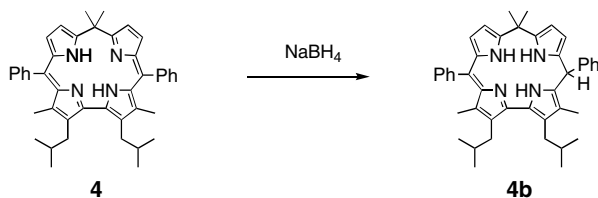
absorption maximum at 586–587 nm. It is noteworthy that the UV–vis feature of **8** (a strong band at 541 nm with shoulders at 459 and 642 nm) is quite different from that of **8a** (a strong band at 435 nm with a weak band at 523 nm), whereas **12** and **12a** show similar UV–vis spectra with a weak band at around 430 nm (433 nm for **12** and 429 nm for **12a**) and a strong band at around 540 nm (538 nm for **12** and 544 nm for **12a**).¹ These UV–vis spectra seem to depend on the conformation of the π -conjugated tetrapyrrolic units and the electronic interaction beyond the bridging sp³ carbons, that is, homoconjugation or transannular π – π interaction. Thus, the UV–vis spectrum converged to that expected for linear oligomers as the ring size increased.

The cyclopolypyrroles larger than cyclododecapyrroles have never been reported in the porphyrinoid chemistry except in our previous work on the expanded porphyrins having 16, 20, and 24 pyrrole units.⁵ To the best of our knowledge, a free base form of isocorrole has never been reported so far either. The isocorrole **4** shows a B-type band at 420 nm and Q-type bands at 620 and 675 nm in the UV–vis spectrum and signals due to the NH protons at 13.6 ppm and pyrrole β -protons at 6.53 and 6.38 ppm in the ¹H NMR. The NaBH₄ reduction of **4**

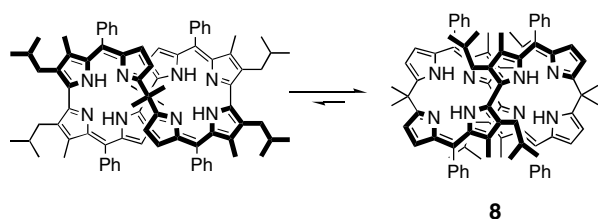
in a mixture of CH_2Cl_2 and ethanol quantitatively gave dihydrocorrole **4b**⁹ (see Scheme 2), which showed three ^1H NMR signals due to the NH protons at 12.6, 8.56, and 8.23 ppm and a singlet due to the *meso*-proton at 5.49 ppm. Therefore, reduction occurred at one *meso*-carbon and one pyrrole nitrogen but not at two *meso*-carbons. Although condensation of **2a** and **3a** in the presence of a large excess amount of $\text{BF}_3 \cdot \text{Et}_2\text{O}$ failed to give isocorrole,¹ it has recently been reported that the acid-catalyzed condensation of dipyrromethane 5,5'-dicarbinol with 2,2'-bipyrrole gave *meso*-triarylcorroles in ca. 10% yield after DDQ oxidation.¹⁰ The latter reaction is closely related to the present reaction, because **2** is a dehydrated form of the dipyrromethane 5,5'-dicarbinol.⁶ Therefore, preparation of **4** in 27.8% yield is remarkable.

It has been reported that the *gem*-dimethyldipyrromethane units are at the crossing point of the figure 8 loop of **8a** on the basis of the X-ray crystallographic analysis and the NOE NMR experiment in solution.¹ A remarkable difference in the UV–vis feature between **8a** and **8** noted above suggests that the loop conformation of **8** is not similar to that of the figure 8 conformation of **8a**. The 2D ROESY NMR spectrum of **8** showed a cross peak between doublets (6.33, 6.18 ppm) due to the β -pyrrole protons and doublets (0.48, 0.47 ppm) due to the methyl protons of the *iso*-butyl group. Therefore, the conformation of **8** is close to the figure 8 loop where the bipyrrole units are at the crossing point as shown in Scheme 3. The conformation of the macrocycle is dependent on the substitution pattern in the macrocycle periphery, especially on the steric bulk of the *meso*-phenyl groups in this case.

The formation of macrocycles using bis(azafulvene) derivative of *gem*-dimethyldipyrromethane and 2,2'-bipyrrole proceeded under neutral conditions to give *gem*-dimethylisocorrole as the smallest member through cyclotetrapyrrole as the largest member. The yield and products distribution were dependent on



Scheme 2. Reduction of isocorrole **4** to dihydrocorrole **4b**.



Scheme 3. The figure 8 loop conformation of cyclooctapyrrole **8**.

the reaction conditions and the substitution pattern of the macrocycle periphery as compared with the result of Vogel and co-workers.¹ Availability of these nano-scale cyclooligomers of different ring sizes with the intervals of 4 pyrrole units is of significance in view of their combinatorial application. Further studies aiming at more selective formation of the giant porphyrinoids as well as their use in the supramolecular chemistry are based on this work.

Acknowledgements

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- Bis(azafulvenyl)methane **2** was obtained from *gem*-dimethyldipyrromethane dicarbinol in more than 90% yield (Setsune, J.; Tanabe, A.; Watanabe, J.; Maeda, S., in preparation): To a mixture of the dicarbinol (0.53 mmol) and 4-dimethylaminopyridine (0.028 mmol) was added a dry ether solution (8 mL) of di-*tert*-butyl dicarbonate (1.57 mmol) under argon. After stirring for 3 h at room temperature, the color of the solution turned bright yellow. Aqueous K_2CO_3 solution (8 mL, 0.02 mol/L) was added to the reaction mixture and it was extracted with diethyl ether. After drying over anhydrous K_2CO_3 , hexane (5 mL) was added to the ether solution and condensed under reduced pressure. A small amount of precipitates formed initially was removed by filtration and the filtrate was evaporated to give **2**. UV–vis (λ_{max} nm (log ϵ) in CH_2Cl_2) 350 (4.61). ^1H NMR (δ -value in CDCl_3) 8.33 (d, 4H, $J = 8.3$ Hz, *o*-Ph-H); 7.43 (t, 4H, $J = 7.3$ Hz, *m*-Ph-H); 7.38 (t, 2H, $J = 7.2$ Hz, *p*-Ph-H); 6.89 (s, 2H, $-\text{CH}=\text{}$); 7.06, 6.62 (d \times 2, 2H \times 2, $J = 4.5$ Hz, β -pyrrole-H); 1.82 (s, 6H, $-\text{CH}_3$). MS (EI) (found/calcd for $\text{C}_{25}\text{H}_{22}\text{N}_2$) 350/350

- (M⁺). Anal. Calcd (%) for C₂₅H₂₂N₂: C, 85.68; H, 6.33; N, 7.99. Found: C, 85.48; H, 6.39; N, 8.17.
- 2,2'-Bipyrrole **3** was prepared in a similar manner to 3,3'-dimethyl-4,4'-diethyl-2,2'-bipyrrole.¹¹ ¹H NMR (δ-value in CDCl₃) 7.73 (br, 2H, NH); 6.54 (d, 2H, *J* = 2.5 Hz, α-py-H); 2.34 (d, 4H, CH₂CHMe); 2.06 (s, 6H, β-py-CH₃); 1.72 (m, 2H, -CH₂CHMe₂); 0.82 (d, 12H, -CH₂CHMe₂). MS (EI) (found/calcd for C₁₈H₂₈N₂) 272/272 (M⁺). Anal. Calcd (%) for C₁₈H₂₈N₂: C, 78.96; H, 10.41; N, 10.19. Found: C, 79.36; H, 10.6; N, 10.28.
 - Typical procedure: A mixture of *gem*-dimethyl-2,2'-bis(6-phenylazafulvenyl)-methane **2** (0.50 mmol), 3,3'-di-*iso*-butyl-4,4'-dimethyl-2,2'-bipyrrole **3** (0.50 mmol), and Zn(II) *p*-*tert*-butylbenzoate (1.0 mmol) in dry CH₂Cl₂ (18 mL) was stirred for 16 h at room temperature under argon. DDQ (1.60 mmol) was then added to the reaction mixture and stirring was continued for an additional 2 h at room temperature. The resulting blue-green solution was passed through Celite that was washed with CH₂Cl₂ and then with acetone. The acetone washings were evaporated and the residue was dissolved in a mixture of CH₂Cl₂ and methanol (20:1). The combined organic solution was shaken with 2% aqueous HClO₄ solution, water, and 0.5% aqueous NaOH solution, sequentially. The organic layer was separated, dried over Na₂SO₄, and evaporated to dryness. The residue was chromatographed on Al₂O₃ (activities II–III) with toluene to give a mixture of expanded isocorroles. A CHCl₃ solution of these expanded isocorroles was injected to HPLC with a combination of GPC columns of JAIGEL-3H (20 × 600 mm, exclusion limit MW 70,000 polystyrene) and JAIGEL-2H (20 × 600 mm, exclusion limit MW 5000 polystyrene) and the separation was performed by using a recycle loop. Compounds **4**, **8**, **12**, **16**, **20**, **24**, **28**, and **32** were separated at the retention time of 190 (at the third cycle), 188 (at the third cycle), 344 (at the sixth cycle), 499 (at the ninth cycle), 486 (at the ninth cycle), 475 (at the ninth cycle), 466 (at the ninth cycle), and 254 (at the fifth cycle) min, respectively, with a CHCl₃ flow rate of 3.8 mL/min. The fraction running faster than **32** was further separated by the combination of GPC columns of JAIGEL-3H and JAIGEL-2.5H (20 × 600 mm, exclusion limit MW 20,000 polystyrene). Compounds **36**, **40**, and **44** were separated at the retention time of 859, 852, and 844 min at the 16th cycle, respectively, with a CHCl₃ flow rate of 3.8 mL/min. The separated fractions corresponding to **4–32** were evaporated. Reprecipitation by slowly evaporating the CH₂Cl₂–methanol solution and filtration afforded pure materials of **4–32**. The yields of **36**, **40**, and **44** were roughly estimated on the basis of their UV–vis absorbances at 500 nm relative to that of **32** during the GPC separation. Isocorrole **4**: UV–vis (λ_{max} nm (log ε) in CH₂Cl₂) 420 (4.77), 620 (3.88), 675 (3.96). ¹H NMR (δ-value in CDCl₃) 13.6 (br, 2H, NH); 7.44–7.48 (m, 10H, *meso*-Ph-H); 6.53, 6.38 (d × 2, 2H × 2, *J* = 4.3 Hz, β-py-H); 2.51 (d, 4H, *J* = 7.5 Hz, -CH₂CHMe); 1.79 (s, 6H, β-py-CH₃); 1.43 (s, 6H, *gem*-CH₃); 1.72–1.77 (m, 2H, -CH₂CHMe); 0.87 (d, 12H, *J* = 6.8 Hz, -CH₂CHMe₂). ESI-MS (found/calcd for C₄₃H₄₆N₄ + H⁺) 619.39/619.38. Anal. Calcd (%) for C₄₃H₄₆N₄: C, 83.45; H, 7.49; N, 9.05. Found: C, 83.60; H, 7.48; N, 9.17. Cyclooctapyrrole **8**: UV–vis (λ_{max} nm (log ε) in CH₂Cl₂) 459 (sh, 4.44), 541 (4.98), 642 (sh, 4.40). ¹H NMR (δ-value in CDCl₃) 12.4 (br, 4H, NH); 7.28–7.46 (m, 20H, *meso*-Ph-H); 6.33, 6.18 (d × 2, 4H × 2, *J* = 4.2 Hz, β-py-H); 2.73, 1.72 (dd × 2, 4H × 2, *J* = 7.3, 14.6 Hz, -CH₂CHMe₂); 1.69 (s, 12H, β-py-CH₃); 1.23 (s, 12H, *gem*-CH₃); 1.18–1.25 (m, 4H, -CH₂CHMe₂); 0.48, 0.47 (d × 2, 12H × 2, *J* = 6.3 Hz, -CH₂CHMe₂). ESI-MS (found/calcd for C₈₆H₉₂N₈ + nH⁺) 1238.75/1238.76 (*n* = 1); 619.89/619.88 (*n* = 2). Anal. Calcd (%) for C₈₆H₉₂N₈: C, 83.45; H, 7.49; N, 9.05. Found: C, 83.29; H, 7.36; N, 9.33. Cyclododecapyrrole **12**: UV–vis (λ_{max} nm (log ε) in CH₂Cl₂) 433 (sh, 4.61), 538 (5.10). ¹H NMR (δ-value in CDCl₃) 12.9 (br, 6H, NH); 7.37 (br, 30H, *meso*-Ph-H); 6.08, 6.02 (br × 2, 6H × 2, β-py-H); 2.98, 1.98 (br × 2, 6H × 2, -CH₂CHMe₂); 1.65 (s, 18H, β-py-CH₃); 1.25 (s, 18H, *gem*-CH₃); 1.36 (m, 6H, -CH₂CHMe₂); 0.63 (br, 36H, -CH₂CHMe₂). ESI-MS (found/calcd for C₁₂₉H₁₃₈N₁₂ + nH⁺) 1857.16/1857.13 (*n* = 1); 928.98/929.07 (*n* = 2); 619.84/619.71 (*n* = 3). Anal. Calcd (%) for C₁₂₉H₁₃₈N₁₂: C, 83.45; H, 7.49; N, 9.05. Found: C, 83.34; H, 7.62; N, 8.81. Cyclohexadecapyrrole **16**: UV–vis (λ_{max} nm (log ε) in CH₂Cl₂) 433 (sh, 4.68), 557 (5.24). ¹H NMR (δ-value in CDCl₃) 12.9 (br, 8H, NH); 7.22–7.30 (m, 40H, *meso*-Ph-H); 6.03 (d, 8H, *J* = 3.7 Hz, β-py-H); 6.10 (br, 8H, β-py-H); 2.17–2.63 (br, 16H, -CH₂CHMe₂); 1.73 (s, 24H, β-py-CH₃); 1.18 (s, 24H, *gem*-CH₃); 1.47–1.53 (m, 8H, -CH₂CHMe₂); 0.68 (d, 48H, *J* = 6.3 Hz, -CH₂CHMe₂). ESI-MS (found/calcd for C₁₇₂H₁₈₄N₁₆ + nH⁺) 2475.00/2475.50 (*n* = 1); 1238.43/1238.76 (*n* = 2); 826.18/826.17 (*n* = 3); 620.03/619.88 (*n* = 4). Anal. Calcd (%) for C₁₇₂H₁₈₄N₁₆: C, 83.45; H, 7.49; N, 9.05. Found: C, 83.63; H, 7.69; N, 9.03.
 - Spectroscopic data of **4b**: UV–vis (λ_{max} nm (log ε) in CH₂Cl₂) 374 (3.93), 425 (3.59), 508 (4.26), 541 (sh, 413). ¹H NMR (δ-value in CDCl₃) 12.6, 8.56, 8.23 (s × 3, 1H × 3, NH); 7.22–7.45 (m, 10H, *meso*-Ph-H); 6.05, 5.91 (d × 2, 1H × 2, *J* = 4.0 Hz, β-py-H); 5.97, 5.73 (t × 2, 1H × 2, *J* = 3.0 Hz, β-py-H); 5.49 (s, 1H, *meso*-H); 2.32, 2.45, 2.49, 2.56 (dd × 4, 1H × 4, *J* = 14.2, 7.5 Hz, -CH₂CHMe₂); 1.70, 1.66 (s × 2, 3H × 2, β-py-CH₃); 1.78, 1.33 (s × 2, 3H × 2, *gem*-CH₃); 1.80–1.85 (m, 2H, -CH₂CHMe₂); 0.91, 0.90, 0.87, 0.78 (d × 4, 3H × 4, *J* = 6.5 Hz, -CH₂CHMe₂). ESI-MS (found/calcd for C₄₃H₄₈N₄ + H⁺) 621.58/621.40.
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